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Genotype distribution in chronic hepatitis C, relationship of genotype with HCV RNA, alpha fetoprotein, alanine aminotransferase and liver fibrosis

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Abstract

In this study, the genotype distribution in chronic hepatitis C (CHC) and its relationship with alanine aminotransferase (ALT), alpha fetoprotein (AFP), HCV RNA and liver fibrosis were compared. HCV RNA was negative in 63 and positive in 69 CHC cases who applied to the infectious diseases outpatient clinic between May 2016 and June 2018 were included in the study. Demographic data, laboratory and pathology results of the cases were evaluated retrospectively. 76 of the cases were male, 56 of them were female. In HCV RNA positive cases, Serum ALT and AFP level were statistically significantly higher than negative ones ($p: 0.04$, $p: 0.001$). Of HCV RNA positive cases, genotype 1b in 42 (60.9%), 3a in 10 (14.5%), 1a in 8 (11.6%), 4d in 5 (7.2%), and 2b in 4 (5.8%) were found. There was no statistically significant difference between genotypes in terms of HCV RNA, ALT and fibrosis. AFP level was statistically significantly higher in genotype 1b than other genotypes. Consistent with studies, the most common genotype was 1b. One of the main results is that there is no significant relationship between genotype difference and ALT, AFP and HCV RNA and fibrosis in CHC cases. Therefore, although these parameters may play a role in chronicity and inflammation, they may not be sufficient in the evaluation of fibrosis. High level of AFP in genotype 1b compared to other genotypes may be useful in determining prognosis. Such studies may be important in epidemiological evaluation of HCV and follow-up of genotype profile changes.

Keywords: Chronic hepatitis C, alanine aminotransferase, alpha fetoprotein, HCV RNA, genotype

Introduction

Hepatitis C virus infection (HCV) is mainly a parenterally transmitted disease with slow progression, but it can progress as an acute infection. While it resolves spontaneously in approximately 20% of the cases, it becomes chronic in 80% of the cases and may result in chronic HCV, liver cirrhosis (LC) and hepatocellular carcinoma (HCC) [1, 2]. The mechanism of liver damage due to HCV has not been fully explained and it is thought that the virus escapes from the immune system as a result of high chronicity rate and high genetic heterogeneity [3]. Hepatitis C virus (HCV) is a single-stranded RNA virus and consists of seven major genotypes (HCV 1-7) with 67 confirmed and 20 transient subtypes, and genotypes 1, 2 and 3 show a wide distribution, 4, 5 and 6 are only seen in certain regions [4].

Genotyping of HCV is important for obtaining epidemiological data, determining prognosis and choosing antiviral therapy. Patients infected with genotype 1b have a higher risk of developing LC and HCC, and this genotype is more resistant to antiviral therapy than other genotypes [5]. The most frequently detected genotype in Turkey are genotype 1 and genotype 3 [6, 7]. In cases with chronic hepatitis C (CHC), HCV RNA and serum alanine aminotransferase (ALT) levels are important markers in showing the degree of viral replication and liver damage, initiating treatment [4].

Prevention of disease transmission and treatment of infected cases are the main strategies for the management of CHC due to the complications that may occur in the future due to the disease and its contagiousness. The disease is almost curable with the introduction of many new and effective drugs in treatment; besides, survival times have significantly increased [8, 9].

In this study, it was aimed to evaluate the genotype distribution and its relationship with ALT, alpha fetoprotein (AFP), viral load (HCV RNA) and liver fibrosis in patients with CHC and to contribute to the diagnosis, determination of prognosis in demonstrating liver damage.

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Materials and Methods

132 anti-HCV (+), HCV RNA positive and negative cases for at least 6 months who admitted to the Infectious Diseases outpatient clinic of Fethi Sekin City Hospital between May 2016 and June 2018 were included in the study. Demographic data of the cases, anti-HCV, HCV RNA and HCV genotype, and liver fine needle aspiration biopsy results were evaluated retrospectively. Anti-HCV enzyme "immunoassay" (ELISA), HCV RNA level Cobas AmpliPrep / Cobas Taqman 48 (Roche Diagnostics, USA), genotype determination was determined by quantitative "real time" HCV Genotype II system (Abbott Laboratories, USA).

Demographic data, ALT, AFP values of HCV RNA positive and negative cases were recorded and comparison was made. In HCV RNA positive cases, liver fine needle aspiration biopsy results were recorded together with the genotype determination. Liver biopsy specimens were scored using the Ishak histological scoring system (fibrosis was evaluated out of 6) (10). Genotype 1a, 1b, 3a were included in the study and a comparison was made between these three genotypes in terms of ALT, AFP, HCV RNA levels and fibrosis.

Patients with a history of hepatotoxic drug and alcohol use, autoimmune diseases, acute hepatitis, LC and HCC, and those aged 18 years or younger were excluded from the study. This study was in accordance with the international ethics committee (Helsinki Declaration) principles and was approved by Firat University Non-Interventional Research Ethics Committee (11.04.2019/06).

Statistical Analysis

Data analysis procedures were carried out using SPSS 22.0 (Chicago, USA) package statistics software. Paired-samples t test was used for the analysis of continuous variables conforming to normal distribution. The analysis of categorical data was made using the chi-square test. One-way test (tukey) was used in multiple comparisons. Numerical data were expressed as mean $\pm$ standard deviation, and categorical data as %. $P < 0.05$ value was accepted as significant in statistical comparisons.

Results

76 of the cases were male (mean age 53.6 ± 3.29), 56 of the cases

were women (mean age 62.9 ± 1.46). In 63 of 132 cases, HCV RNA was negative, 69 of them were HCV RNA positive and HCV RNA levels were between 3553-34.243.424. 990 IU / mL. The demographic characteristics of HCV RNA negative and positive cases are summarized in Table 1.

Table 1. Demographic characteristics of HCV RNA negative and positive chronic hepatitis C cases

	HCV RNA negatif	HCV RNA pozitif	p value
	Avarage $\pm$ SD	Avarage $\pm$ SD	
Age	52.7 $\pm$ 2.40	58.3 $\pm$ 2.04	0.48
Gender (F/M)	27/30	21/43	0.42
AST(IU/L)	23.28 $\pm$ 3.96	32.77 $\pm$ 3.67	0.05
ALT(IU/L)	22.86 $\pm$ 1.93	38.40 $\pm$ 4.41	0.04
AFP	2.67 $\pm$ 0.20	7.03 $\pm$ 1.54	0.001

Alanine aminotransferase; AFP: Alpha feto protein; AST: Aspartate aminotransferase; M: Male; F: Female

No statistically significant difference was found between HCV RNA negative and positive cases in terms of age and gender ($p > 0.05$). However, ALT and AFP levels were higher in HCV positive group than negatives and this was statistically significant (38.40 ± 4.41 IU / ML, 7.03 ± 1.54 μ g / L, respectively, $p: 0.04$, $p: 0.001$). There was no statistically significant difference between serum ALT, AFP and HCV RNA levels and fibrosis values in HCV RNA positive cases ($p: 0.383$, $p: 0.211$, $p: 0.324$, respectively).

Of the cases, genotype 1b in 42 (60.9%), genotype 3a in 10 (14.5%), genotype 1a in 8 (11.6%), genotype 4d in 5 (7.2%), genotype 2b in 4 (5.8%) were detected. The mean age of cases with genotype 1b was higher than 1a and 3a and this was statistically significant ($p: 0.001$, $p: 0.003$, respectively). There was no significant relationship between genotypes in terms of gender ($p: 0.218$). In Table 2, ALT, AFP, HCV RNA and fibrosis values are given according to genotypes. There was no statistically significant difference between genotypes in terms of HCV RNA, ALT and fibrosis. Alpha feto protein level was higher than 1a and 3a in genotype 1b and this was statistically significant ($p: 0.03$, $p: 0.04$). Table 3 shows p values of ALT, AFP, HCV RNA and fibrosis distribution according to genotypes.

Table 2. ALT, AFP, HCV RNA and fibrosis values by genotypes

Genotype	Age	ALT	AFP	HCV RNA	Fibrosis
1a	35.3 $\pm$ 4.62	32.14 $\pm$ 12.9	3.5 $\pm$ 2.8	5602700 $\pm$ 2947510	2.5 $\pm$ 1.7
1b	62.1 $\pm$ 2.26	32.42 $\pm$ 5.54	6.9 $\pm$ 11.6	929950.10 $\pm$ 5628840.10 ³	3.4 $\pm$ 2.0
3a	41.6 $\pm$ 7.93	44.62 $\pm$ 12.3	2.3 $\pm$ 1.6	5035900 $\pm$ 7907680	1.7 $\pm$ 0.6

Alanine amino transferase; AFP: Alfa feto protein

Table 3. P values between genotypes and ALT, AFP, HCV RNA and fibrosis

p values	ALT	AFP	HCV RNA	Fibrosis
p1	0.23	0.03	0.24	0.26
p2	0.24	0.24	0.24	0.66
p3	0.24	0.04	0.22	0.14

p1: Genotype 1a-1b; p2: Genotype 1a-3a; p3: Genotype 1b-3a; ALT: Alanine amino transferase; AFP: Alfa feto protein

Genotypes 2b and 4d were not evaluated because they were seen in a small number of cases.

Discussion

Hepatitis C virus infection has an important place among liver diseases due to its high rate of chronicity and transmission routes [11]. In this study, genotype 1b in 42 (60.9%) of the HCV RNA positive cases, and genotype 3a in 10 (14.5%) and genotype 1a in 8 (11.6%) were found. There was no significant difference between genotypes in terms of HCV RNA, ALT and fibrosis ($p>0.05$). Alpha feto protein level was statistically significantly higher in genotype 1b than 1a and 3a ($p: 0.03$, $p: 0.04$, respectively).

The frequency of HCV genotypes in a population varies depending on the age of infection and the route of transmission [5]. In one study, it was observed that the majority of CHC cases admitted to the hospital were among 50 years or older [12]. In another study, Aydemir et al. [13] evaluated 97,527 patients retrospectively and found that the highest positivity value of anti-HCV result was among those of 50 years and above.

Harman et al. [12] found that 75% of the cases were in those of 50 years old and above. 104 (65%) of the cases were female and 56 (35%) were male. Similarly, in a study assessing the epidemiology of HCV, HCV prevalence in all age groups were seen to be higher among women than men in Turkey [14, 15]. Küçüköztepe et al. [16] reported that the average age of patients infected with genotype 1 was statistically significantly higher than those infected with genotype 2 and 3, but there was no significant relationship between genotypes in terms of gender ($p: 0.218$).

In another study, according to genotypes, no statistically significant difference was found in terms of age ($p = 0.431$) and gender ($p = 0.057$) distributions [17]. In this study, also in accordance with the others, 52.3% of CHC cases were among 50 years and older. The mean age of the cases with genotype 1b was statistically significantly higher than 1a and 3a ($p: 0.001$, $p: 0.003$, respectively). However, there was no significant relationship between genotypes in terms of gender ($p: 0.218$).

There are studies stating that serum ALT levels are important in determining liver damage and may be a guide in liver biopsy and treatment decisions [18]. Al Swaff [19] found that there is a relationship between grade 1 and grade 4 liver fibrosis and ALT levels, and higher HCV RNA levels are found in grade 3 liver fibrosis in patients with CHC. In another study, no clinically appropriate relationship was found between ALT levels and liver fibrosis [3]. In the studies of Küçüköztepe et al. [16] including 52 cases, genotype 1b in 76.9% of the cases, genotype 3a in 9.6%, genotype 4e in 5.7%, genotype 2a / 2c in 3.8%, genotype 4 and 1a in 1.9% were found. Again, in one study, no statistically significant difference was found between genotypes in terms of HCV RNA levels and ALT levels ($p> 0.05$) [17]. In this study, there was no significant difference between genotypes between HCV RNA and serum ALT levels.

Due to the genetic diversity of the HCV, it can escape from the immune system and this may cause resistance in treatment and therefore, determination of the genotype can guide the determination of the prognosis and the appropriate treatment

regimen [5]. Genotype 1b is associated with poor prognosis and is more resistant than other genotypes [20]. In various studies [3], the relationship between the degree of liver damage and serum ALT, HCV RNA levels and HCV genotype in cases with CHC has been investigated, but inconsistent results have been reported [3]. Shahid et al. [2] found that HCV RNA titers are associated with the degree of fibrosis in CHC genotype 3a.

In their study including 500 HCV RNA positive patients, Kabakçı et al. [21] showed that the most frequent genotype was 1b (93.5%) and the second most common genotype was 1a (6.7%). In the study carried out by Öztürk et al in Antakya [22], they reported the frequency of type 1a as (0.31%), 1b as (86.73%), 2 as (9.26%), 3 as (0.93%) and 4 as (2.78%). Of 308 patients with CHC, genotype 1 in 271 (88%), genotype 3 in 15 (4.9%), mix genotype in 9 (2.9%), genotype 2 (2.6%) 8 and genotype 4 (1.6%) in 5 were detected by Uzun et al. [23]. Chakravarti et al. [24] found 63.39% type 3, 30.98% type 1, 5.63% type 2 in their study on 71 cases. Gower et al. [25], analyzing 2,320 studies from 98 countries, examined the HCV genotype distribution and genotype 1 constitutes 46% of HCV infections in adults (22% type 1b) and others are distributed as genotype 3 (22%), genotype 2 (13%), genotype 4 (13%), genotype 6 (2%) and genotype 5 (1%).

In this study, in accordance with the studies, the most common genotype was genotype 1b with a rate of 60.9%. No statistically significant difference was found between genotypes in terms of fibrosis scores. However, AFP level was statistically significantly higher in genotype 1b than 1a and 3a ($p: 0.03$, $p: 0.04$, respectively).

The exclusion of patients with hepatocellular carcinoma and advanced stage fibrosis and the relatively small sample size were the limitations of the study.

In conclusion, determination of genotype distribution can be important in determining prognosis and selection of antiviral therapy. In this study, consistent with the studies, the most common genotype was found to be 1b. One of the main results of this study is that there is no significant relationship between genotype difference and ALT, AFP and HCV RNA and fibrosis in CHC patients. Therefore, although these parameters may play a role in chronicity and inflammation, they may not be sufficient in the evaluation of fibrosis. High level of AFP in genotype 1b compared to other genotypes may be useful in determining prognosis. Such studies may be important in epidemiological evaluation of HCVs and follow-up of genotype profile changes. For the evaluation of treatment results and the relationship with fibrosis, there is a need to study larger patient populations, including cases of advanced fibrosis.

Conflict of interests

The authors declare that they have no competing interests.

Financial Disclosure

All authors declare no financial support.

Ethical approval

This study was in accordance with the international ethics committee (Helsinki Declaration) principles and was approved by Firat University Non-Interventional Research Ethics Committee (11.04.2019/06).

References

1. Kuo MT, Hu TH, Lu SN, et al. Neutrophil-to-lymphocyte ratio as a predictor of response to peginterferon plus ribavirin therapy for chronic hepatitis C. *Dis Markers*. 2014;2014:1-17.
2. Shahid M, Idrees M, Nasir B, et al. Correlation of biochemical markers and HCV RNA titers with fibrosis stages and grades in chronic HCV-3a patients. *Eur J Gastroenterol Hepatol*. 2014;26:788-94.
3. Nar R, Sezgin MF. The relationship between the serum RNA titers of hepatitis C Virus and Biochemical Parameters in Chronic Hepatitis C Patients. *Viral Hepatitis J*. 2016;22:28-33.
4. Chen Y, Yu C, Yin X, et al. Hepatitis C virus genotypes and subtypes circulating in Mainland China *Emerging Microbes & Infections*. 2017;6:95.
5. Zeytinli ÜO, Yücel FM, Dinçer ŞD, et al. Distribution of hepatitis C virus genotypes in the region of istanbul northern anatolian association of public hospitals. *Viral Hepatitis J*. 2017;23:10-3.
6. Caliskan A, Kirisci O, Ozkaya E, et al. Distribution and predominance of genotype 3 in hepatitis C virus carriers in the province of Kahramanmaraş, Turkey. *Hepat Mon*. 2015;15:25142.
7. Altuğlu I, Sertoş R, Aksoy A, et al. Possible transmission risks and genotype distribution of hepatitis C virus infection in Western Turkey. *Turk J Gastroenterol*. 2013;24:349-55.
8. Marinho RT, Vitor S, Velosa J. Benefits of curing hepatitis C infection. *J Gastrointestin Liver Dis*. 2014;23:85-90.
9. Saludes V, González V, Planas R, et al. Tools for the diagnosis of hepatitis C virus infection and hepatic fibrosis staging. *World J Gastroenterol*. 2014;20:3431-42.
10. Ishak K, Baptista A, Bianchi L, et al. Histological grading and staging of chronic hepatitis. *J Hepatol*. 1995;22:696-9.
11. Li HC, Lo SY. Hepatitis C virus: Virology, diagnosis and treatment. *World J Hepatol*. 2015;7:1377-89.
12. Harman R, Günel Ö, Özger S. Gaziantep bölgesindeki kronik hepatit C hastalarında hepatit C virüsü genotip dağılımı. *Klinik Dergisi*. 2017;30:68-70.
13. Aydemir Ö, Demiray T, Koroğlu M, et al. Hepatitis C prevalence in different age groups; people over 50 years of age may receive one-time testing for anti-HCV. *Viral Hepatitis J*. 2015;21:40-3.
14. Bruggmann P, Berg T, Øvrehus AL, et al. Historical epidemiology of hepatitis C virus (HCV) in selected countries. *J Viral Hepat*. 2014; 21(Suppl. 1):5-33.
15. Tozun N, Ozdoğan O, Cakaloglu Y, et al. Seroprevalance of hepatitis B and C virus infections and risk factors in Turkey: a fieldwork TURHEP study. *Clin Microbiol Infect*. 2015;21:1020-6.
16. Küçüköztas MF, Özgüneş N, Yazıcı S. Kronik hepatit C'li hastalarda hepatit C virüsü (HCV) genotipleri ile alanin aminotransferaz ve HCV RNA düzeyleri arasındaki ilişkinin araştırılması. *Mikrobiyol Bul*. 2010;44:111-5.
17. Çizmeci Z. Kronik hepatit C enfeksiyonlu hastalarda hepatit C virüs genotiplerinin dağılımı. *Türk Mikrobiyol Cem Derg*. 2016;46:27-32.
18. Kirişçi Ö, Çalışkan A, Koçtürk SA, et al. Kahramanmaraş ili hepatit C virüs ile enfekte bireylerde genotip dağılımı ve genotipin HCV-RNA yükü ve ALT-AST ilişkisi. *Viral Hepatit Dergisi*. 2013;19:67-70.
19. Al Swaff R. Correlation between alanine aminotransferase level, HCV-RNA titer and fibrosis stage in chronic HCV genotype 4 infection. *The Egyptian Journal of Medical Human Genetics*. 2012;13:207-12.
20. Us T, Kaşifoğlu N, Aslan FG, et al. The distribution of hepatitis C virus genotypes of patients with chronic hepatitis c infection in The Eskisehir Region of Turkey. *J Clin Anal Med*. 2017;8:88-91.
21. Kabakçı AG, Karataylı SC, Karataylı E, et al. Hepatitis C virus genotype distribution in Turkey remains unchanged after a decade: performance of phylogenetic analysis of the NS5B, E1, and 5'UTR regions in genotyping efficiency. *Turk J Gastroenterol*. 2014;25:405-10.
22. Oztürk AB, Doğan UB, Oztürk NA, et al. Hepatitis C virus geno5 types in Adana and Antakya regions of Turkey. *Turk J Med Sci*. 2014;44:661-5.
23. Uzun B, Şener AG, Güngör S, et al. Distribution of Hepatitis C virus genotypes in western Turkey: experience of four years. *Acta Medica Mediterranea*. 2014;30:1109-13.
24. Chakravarti A, Dogra G, Verma V, et al. Distribution pattern of HCV genotypes & its association with viral load. *Indian J Med Res*. 2011;133:326-31.
25. Gower E, Estes C, Blach S, et al. Global epidemiology and genotype distribution of the hepatitis C virus infection. *J Hepatol*. 2014;61:45-57.